

Monitoring microbial surface loads, how is it done, what do we find?

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What do we mean by it?

Monitoring the presence of microorganisms on environmental surfaces within the hospital setting

- **Why** should we do it?
- **When** should we do it?
- **How** do we do it?
- **Where** should we look?
- **What do we find** when we do look?

Why should you do it?

- Environmental surfaces play an **important role in the dissemination** of microorganisms,¹
- Detect presence and movement of **clinically-relevant microorganisms** on environmental surfaces in the hospital setting
- Support existing **clinical surveillance**
- Allow for **targeted** infection prevention and control interventions

When would you do it?

Routine monitoring

- Regular sampling of environmental surfaces to determine microbial loading
- High-risk surfaces

Responsive monitoring

- In response to a clinical event (e.g. for outbreak monitoring or unexpected transmissions)
- Variety of relevant surfaces

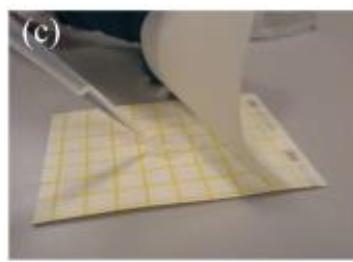
How would you do it?

Sample Collection

Depends on **downstream processing requirements**

Direct

- RODAC Plates
- Dipslides
- Petrifilms



Indirect

- Swabs
- Sponges
- Wipes



How would you do it?

Sample Processing

Depends on **type** of sample and **aim** of sampling

Culture based

- Selective vs non-selective media
- Species level identification
- Antibiotic resistance profiles



Molecular

- qPCR – specific target
- Sequencing



Where would you do it?

Important to consider a **range of sites** – not just the ‘classic’ high-risk environmental surfaces:

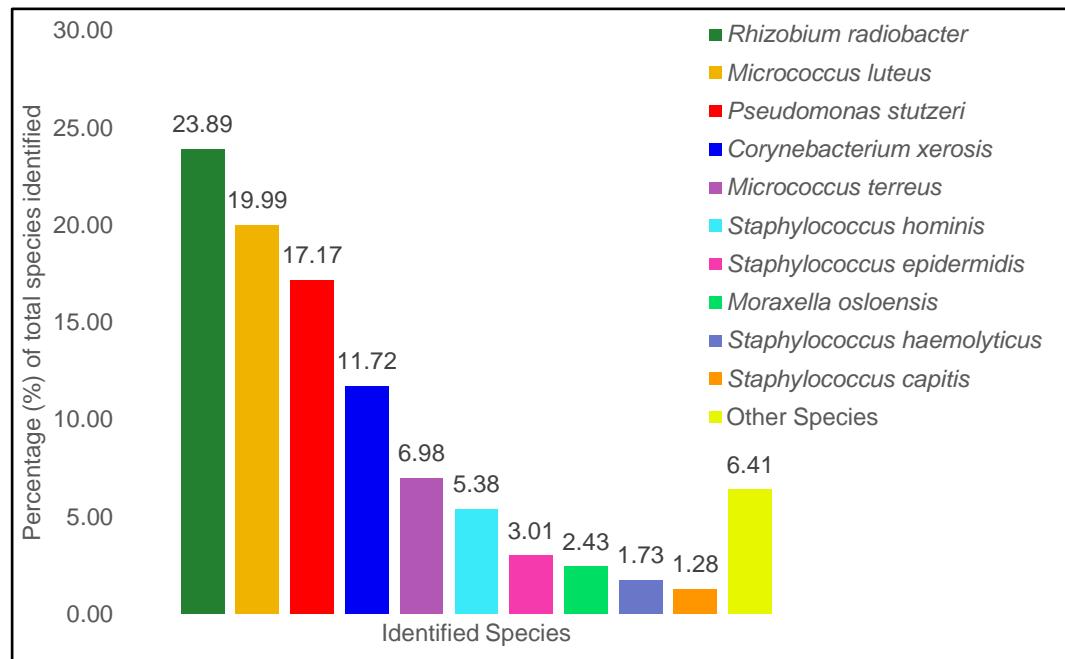
- Non-clinical areas
- Low-touch sites
- Communal touch sites
- Sinks and other water outlets



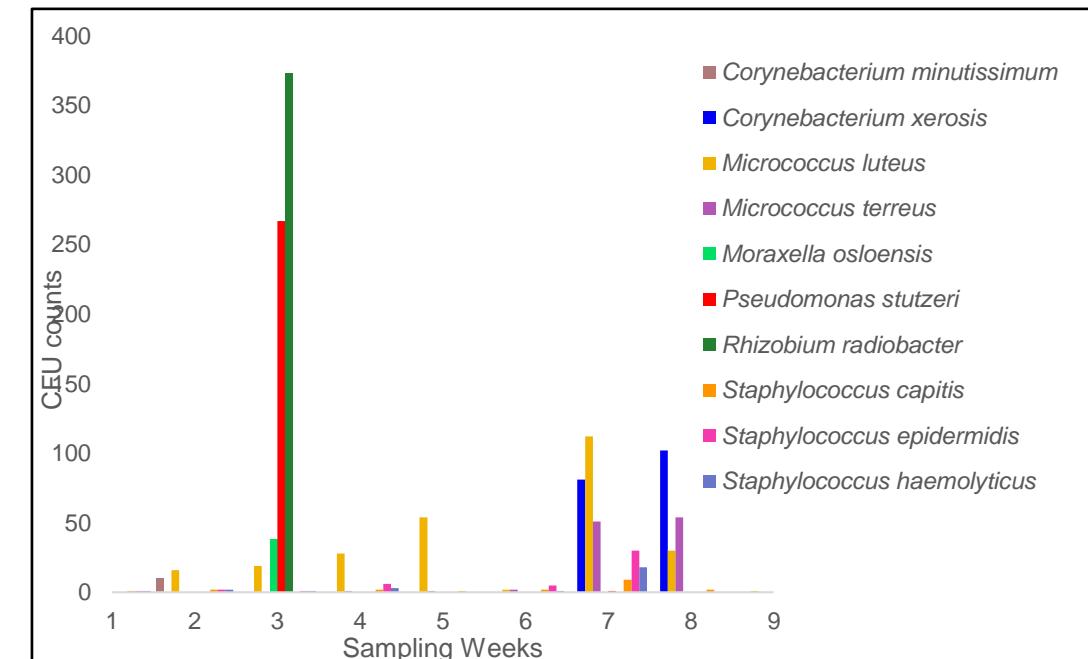
What do we find?

Communal Touch Surfaces

- 22 environmental samples were taken weekly over a **9 week period** in the bone marrow transplant unit at Great Ormond Street Hospital
- Samples were cultured on **Columbia blood agar**
- Aerobic colony counts per 100cm² were established and species were identified by **MALDI-ToF Mass Spectrometry**



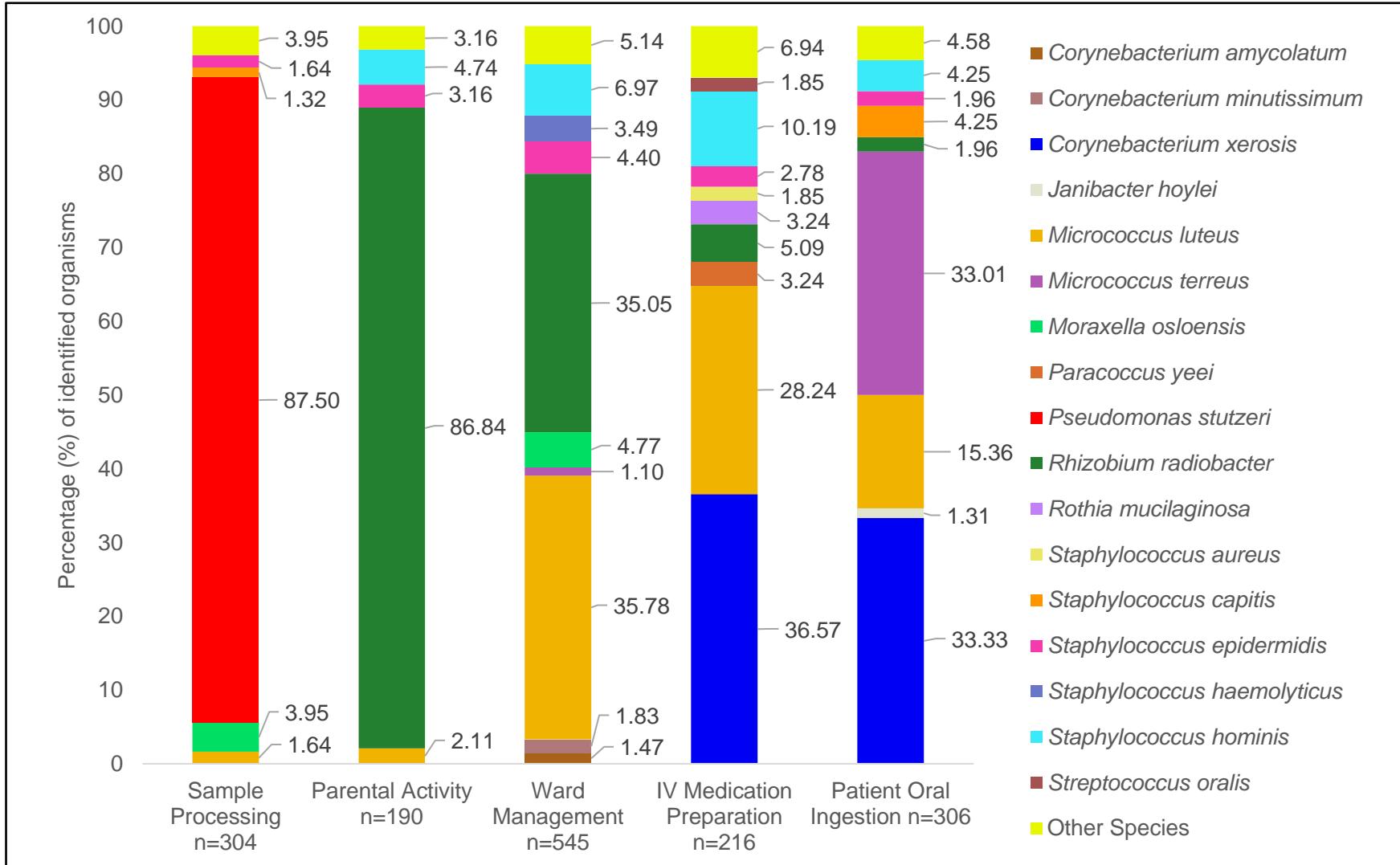
Proportions of all identified species. A total of 42 distinct species were identified. Only species comprising a percentage of the total organisms identified of over 1% are shown.



Distribution of frequently recovered species across the duration of the sampling study. Counts of frequently identified species are plotted against the sampling weeks they were recovered on. Species identified ≥ 10 times are included.

What do we find?

Communal Touch Surfaces



Distribution of identified organisms on sample sites grouped by activity. Sample sites are grouped by activity. All species which comprise $\geq 1.00\%$ of the total organisms present are displayed. Species which comprise $\leq 1.00\%$ are grouped as “Other Species”.

What do we find? Sink Surfaces

Why look at sink surfaces?

- **Unique** environment
- Mechanism of **dispersal**
- **Complex** environment to sample



Aim: Determine the effect patients have on the sink microbiota, and to characterise the sink *Pseudomonas* population in terms of drug resistance and presence of virulence factors

What do we find? Sink Surfaces

- **Non-outbreak** situation
- **Post-operative and Respiratory** wards
- **Flocked swabs** with charcoal transport media
- Sink surface and plug hole
- Plated onto **chromogenic** agar
- Identified by **MALDI-ToF MS**



Examples of sinks sampled in this study

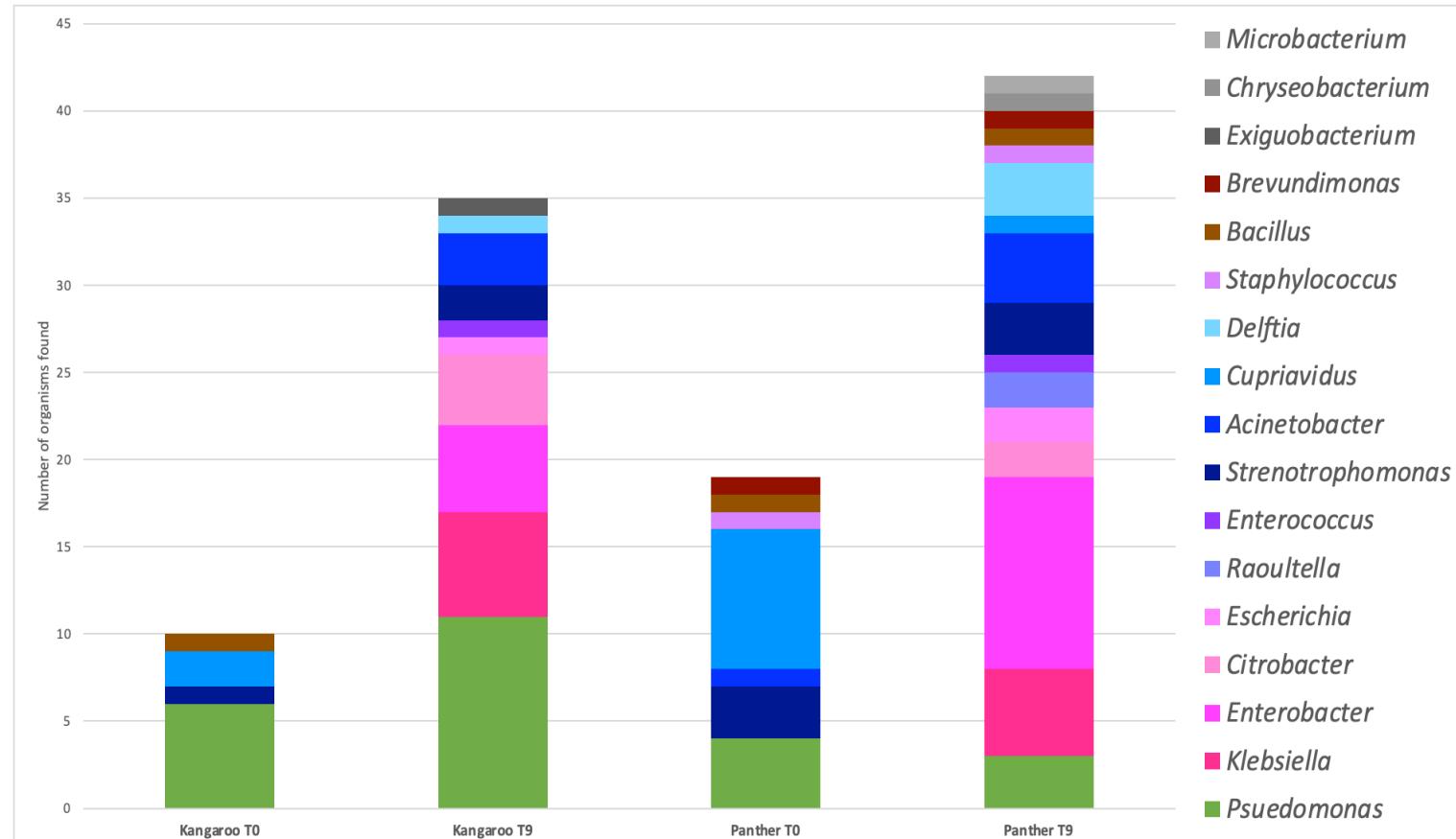
What do we find? Sink Surfaces

Differences after patients

- **Increased amounts** and diversity after patients arrive on both wards
- Move from water and environmental organisms to **human commensals**

Differences between wards

- ***Pseudomonas*** spp. most common after patients introduced in Respiratory ward
- ***Enterobacter*** spp. most common in Post-operative ward
- ***P. aeruginosa*** increases after patients arrive in the Respiratory but not Post-operative ward



Organisms recovered from sinks in Respiratory and Post-operative wards before and after wards open to patients.

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